

# Texas A&M HPRC Short Course Series

## Drug Docking with Schrodinger

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**DIVISION OF RESEARCH**  
TEXAS A & M UNIVERSITY

# Outline

10:00 -10:20 Intro to Molecular Modeling in Drug Discovery

10:20 -11:10 Hands-on Session 1 – Structure Preparation with Maestro

11:10-11:30 Basics of Structural Based Virtual Screening

11:30-12:15 Hand-on Session 2 – Docking with Glide

12:15-12:30 Covalent Docking with Covdock & Wrap-up

# A Tough Road of Drug Discovery

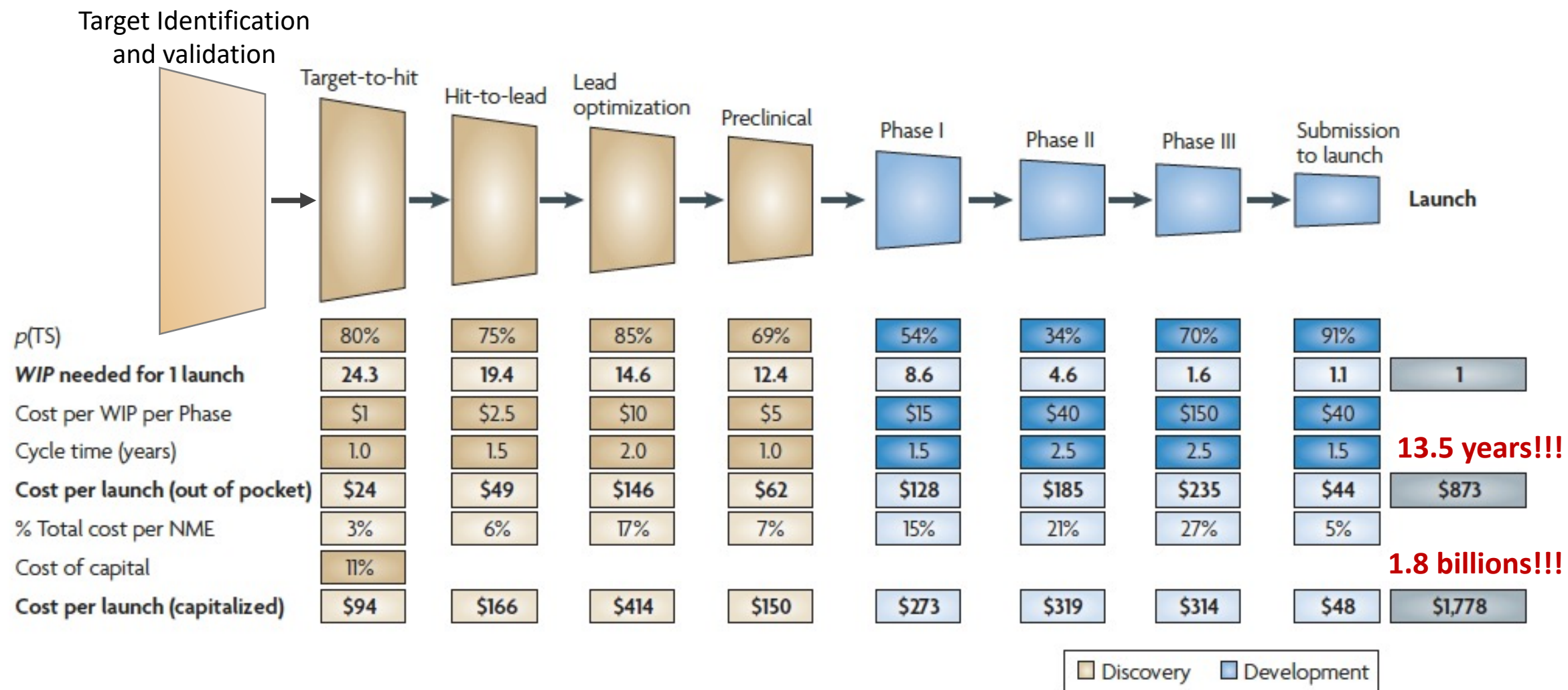
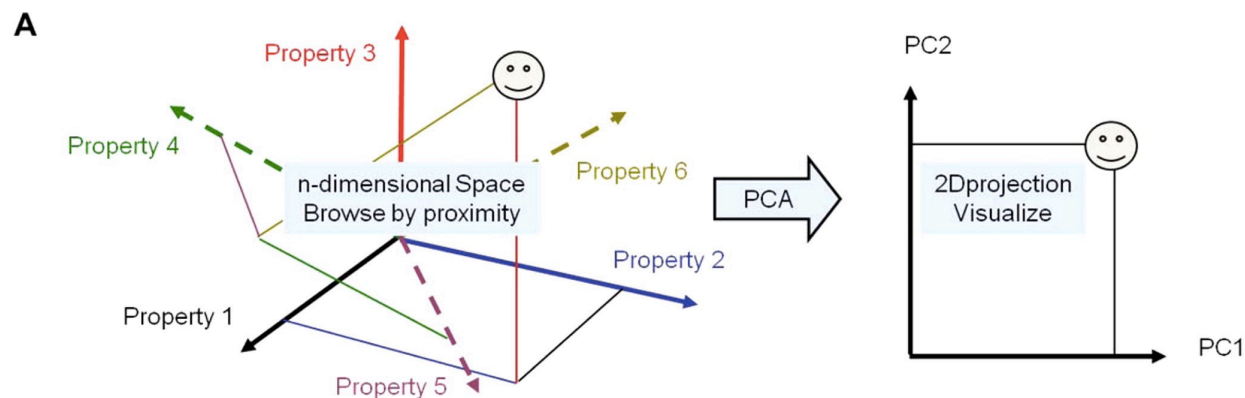
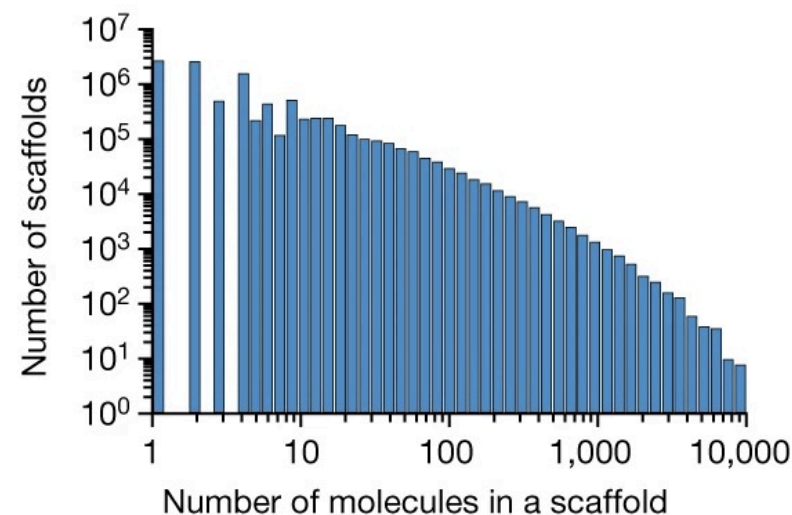
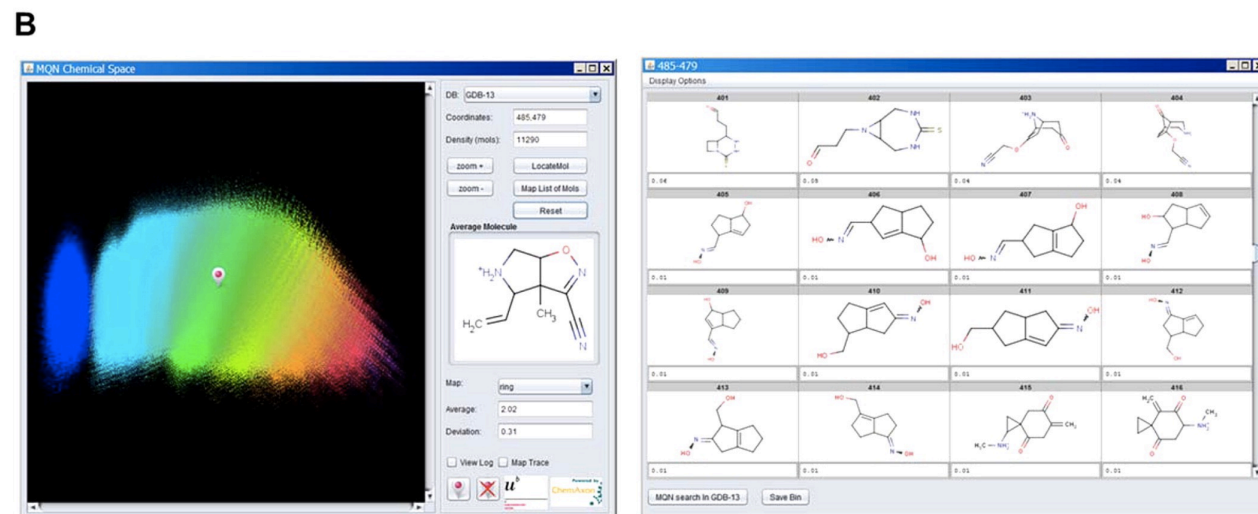


Figure adapted from Paul SM et al., Nat. Rev. Drug Discov., 2010, 9, 203-214

# CADD - Larger chemical space, new hits



- Chemical diversity (scaffolds) increases with large chemical space searching
- As screening decks expand there will be more tighter binders that could be found



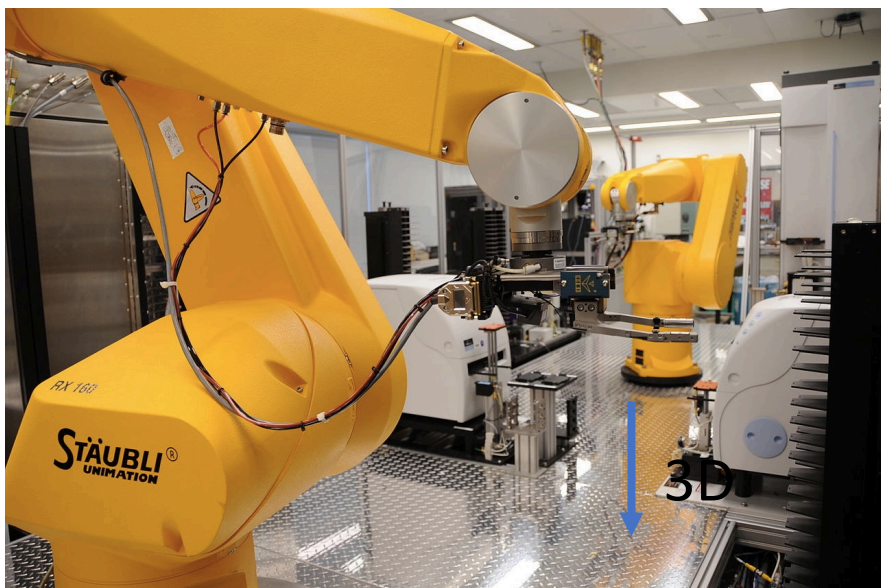
Acc. Chem. Res. 2015, 48, 3, 722–730

Nature 2019, 566, 224–229

# Methods for Hit Identification

**Hit:** A small molecule that is known to bind to a target in drug discovery.

## High Throughput Screening (HTS)



[https://en.wikipedia.org/wiki/High-throughput\\_screening](https://en.wikipedia.org/wiki/High-throughput_screening)

## High Throughput Virtual Screening (HTVS)



**ligand-based**

- 2D Fingerprint searching
- 2D/3D pharmacophore
- 2D/3D QSAR Models

**structure-based**

- 3D
- **Docking**
  - pharmacophore screening



# Virtual Screening

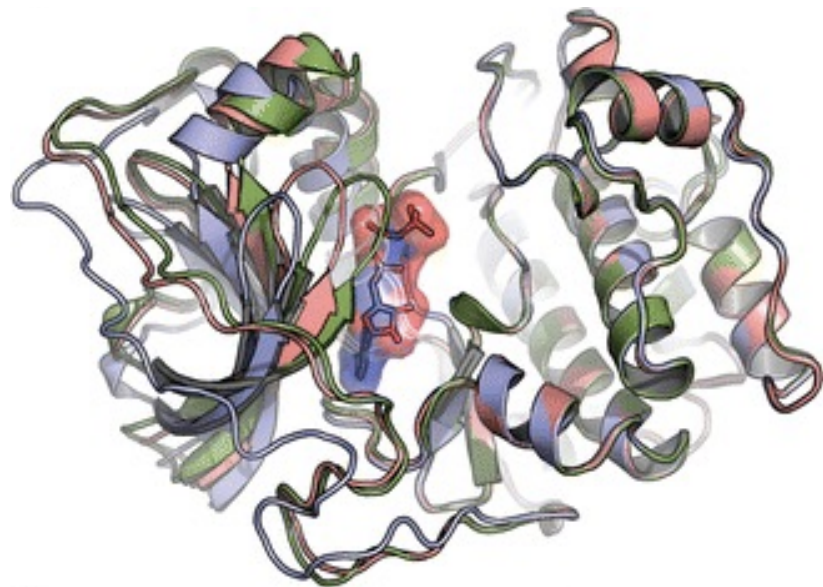


Figure from *J. Am. Chem. Soc.* 2013, 135, 15, 5819–5827

## Structure-based Drug Design

- known target structure
- known ligand binding site
- (optional) bound ligands/hits

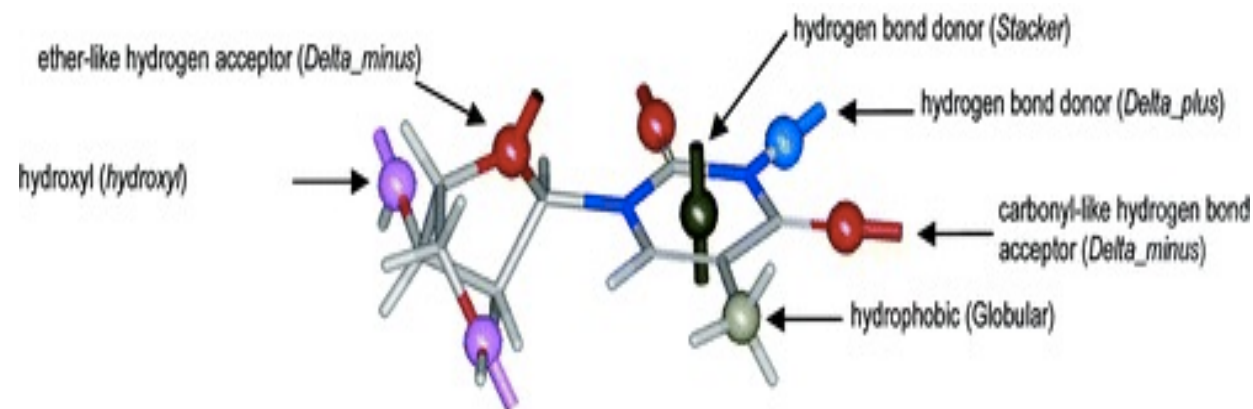


Figure from *J. Chem. Inf. Model.* 2007, 47, 3, 1097–1110

## Ligand-based Drug Design

- known hits
- (optional) active conformation

# Protein Target

- Crystal structure

[RCSB Protein Data Bank \(PDB\)](https://www.rcsb.org/)

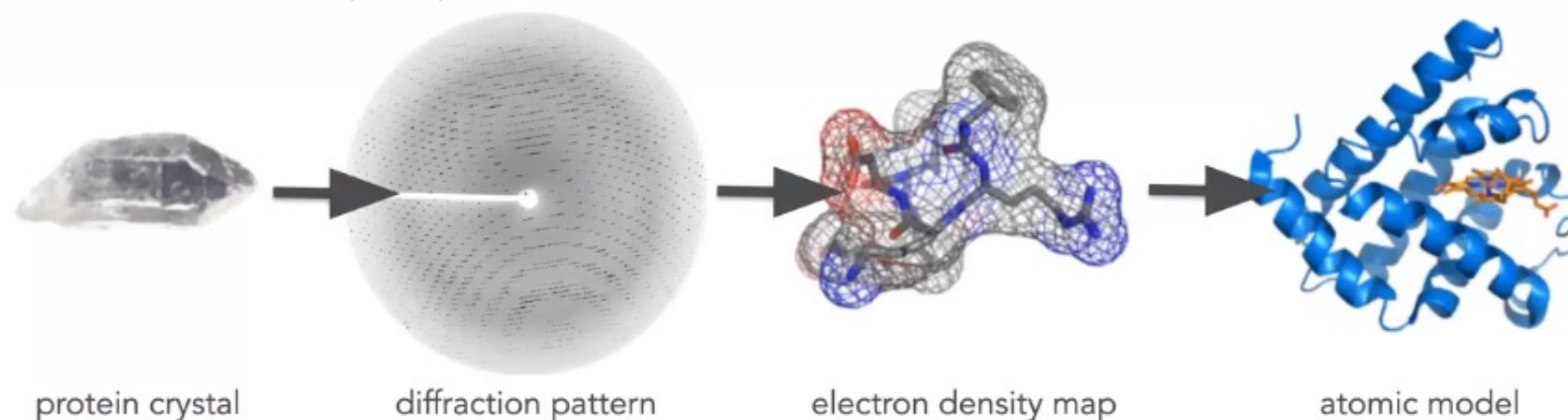
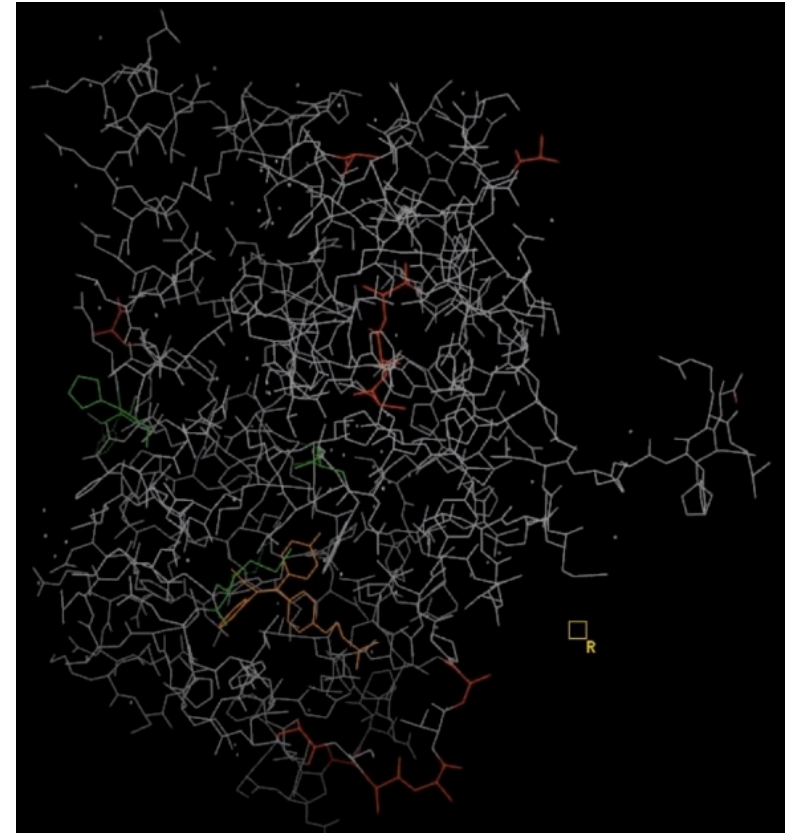


Figure from <https://www.schrodinger.com/webinars/archives/1248/virtual-screening/469153>

- NMR
- Homology Model
- Cryoelectron Microscopy (cryo-EM)

# Protein Preparation

- Typical PDB structure is not suitable for immediate use
  - it typically contains heavy atoms, co-crystallized ligand, water molecules, metal ions, cofactors, ...
  - may be multimeric, need to be reduced to a single unit
  - limited resolution, eg. it's difficult to distinguish carbonyl oxygen and secondary amine nitrogen's of amide
  - may have incorrect bond orders, assignment of charge state, orientation of groups



Color protein with **PDB conversion Status**

Gray: standard residue connect by standard

Red: standard residue with missing atoms

orange: nonstandard residues, HET groups

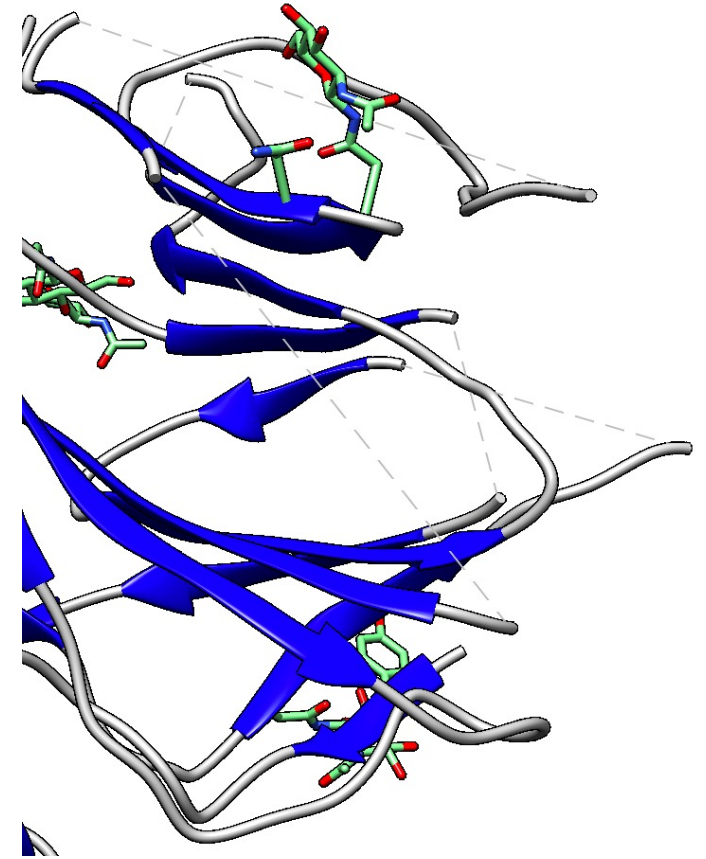
green: residue with an alternate conformation

7



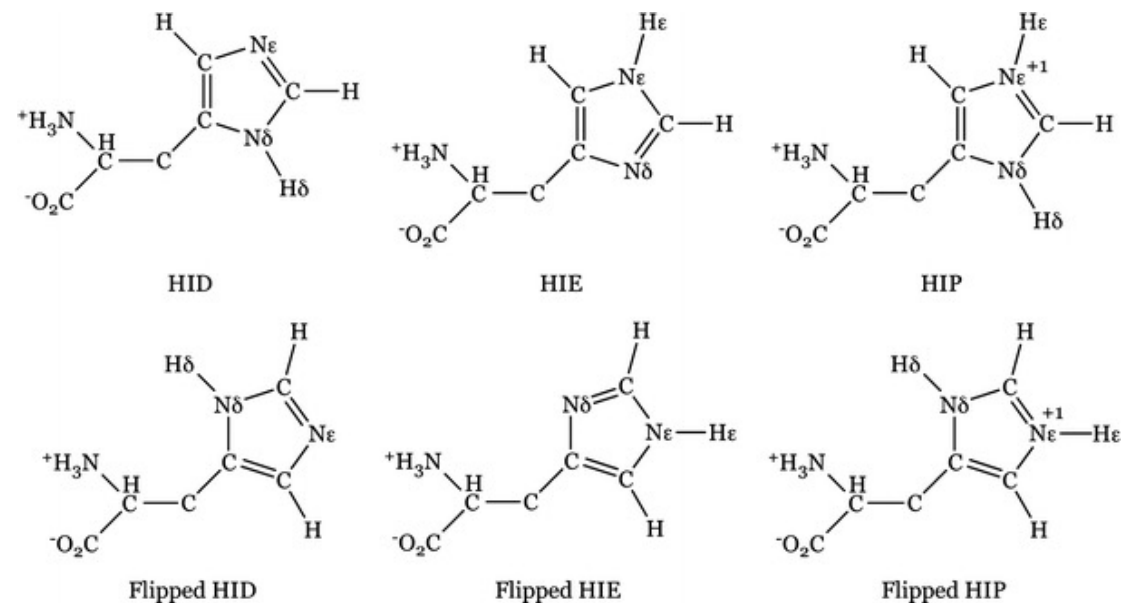
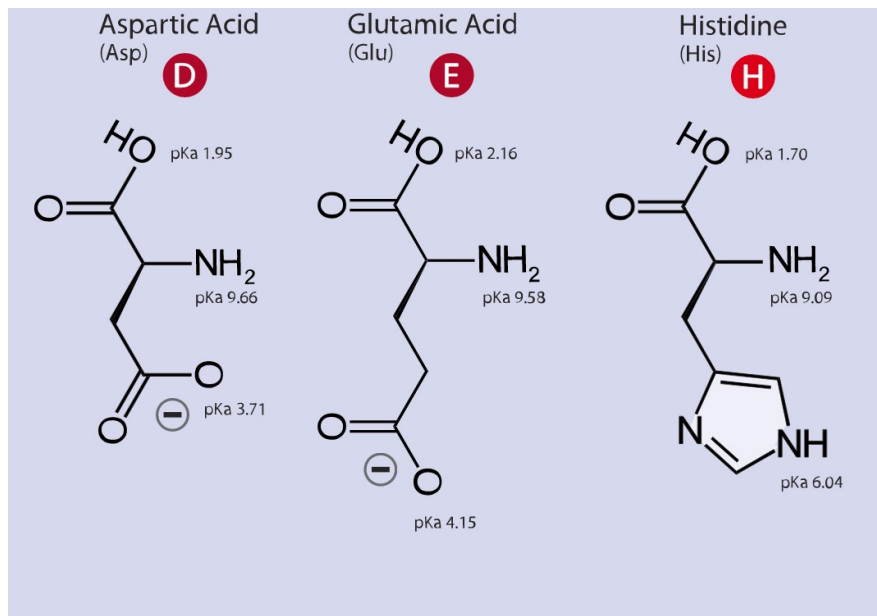
# Protein Preparation – Missing atoms

- Missing atoms
  - Hydrogens are not included
  - Entire side chains may be missing
  - There are a number of utilities to fill in missing atoms/sidechain
- Missing segments
  - More complicated to fix
  - Normally requires homology modeling to obtain reasonable results if more than a few residues are missing



# Protein Preparation – Protonation states

- ASP, GLU and HIS



Adapted from [https://commons.wikimedia.org/wiki/File:Amino\\_Acids.svg](https://commons.wikimedia.org/wiki/File:Amino_Acids.svg)  
Dancojocari / CC BY-SA (<https://creativecommons.org/licenses/by-sa/3.0>)

<https://link.springer.com/article/10.1007/s10822-013-9643-9>

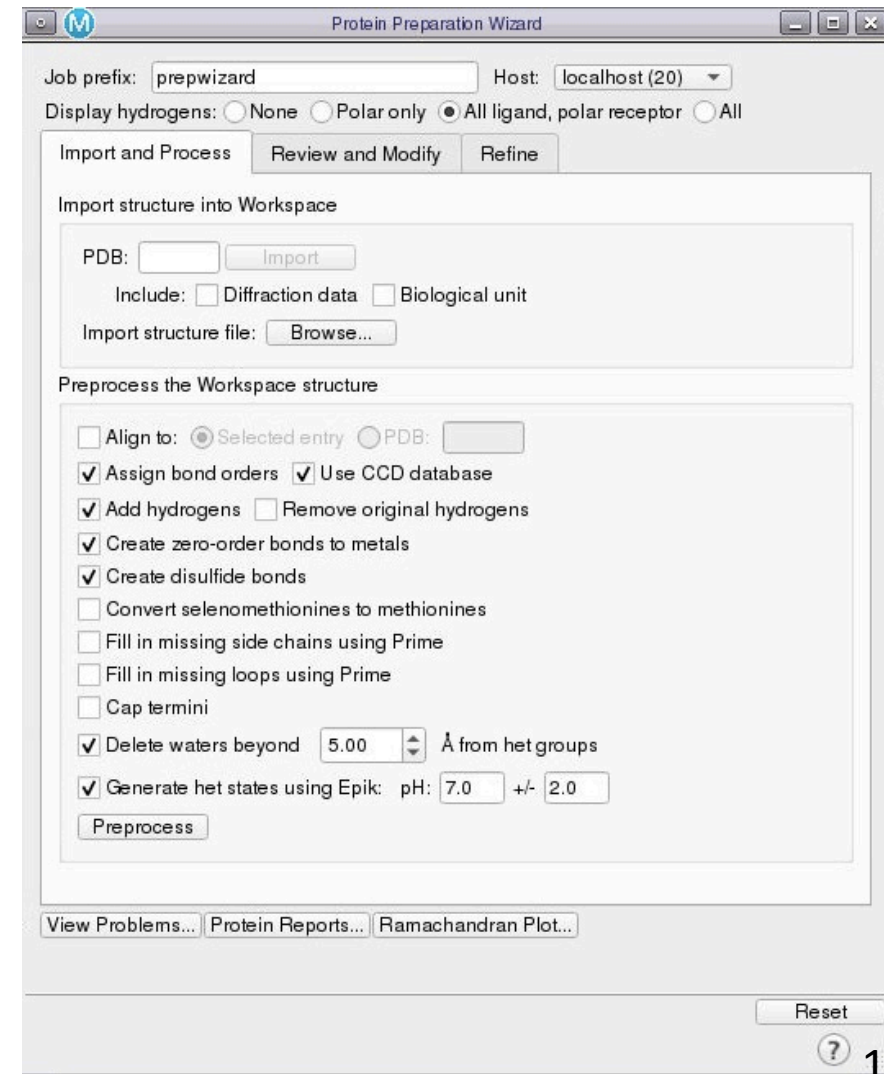
# Protein Preparation Wizard in Schrodinger

- **Import and Process Tab:** fix common problems
  - Protonation
  - Missing Side Chain
  - Missing Loops
- **Review and Modify:** Remove Unwanted Molecule
  - counterions, artifact of crystallography, waters
  - Biologically relevant
- **Refine:** Optimize your structure
  - Hydrogen bonded optimization
  - Remove waters?
  - Restrained Minimization
- **View Problems...**

The screenshot shows the 'Protein Preparation Wizard' window. At the top, the 'Job prefix' is 'prepwizard' and the 'Host' is 'localhost (20)'. Below this, 'Display hydrogens' is set to 'All ligand, polar receptor'. The 'Import and Process' tab is selected, showing options to 'Import structure into Workspace' with a 'PDB' field and 'Import' button, and 'Import structure file' with a 'Browse...' button. The 'Preprocess the Workspace structure' section contains several checkboxes: 'Align to' (Selected entry), 'Assign bond orders' (checked), 'Use CCD database' (checked), 'Add hydrogens' (checked), 'Remove original hydrogens' (unchecked), 'Create zero-order bonds to metals' (checked), 'Create disulfide bonds' (checked), 'Convert selenomethionines to methionines' (unchecked), 'Fill in missing side chains using Prime' (unchecked), 'Fill in missing loops using Prime' (unchecked), 'Cap termini' (unchecked), 'Delete waters beyond 5.00 Å from het groups' (checked), and 'Generate het states using Epik' (checked) with pH 7.0 and +/- 2.0. A 'Preprocess' button is at the bottom of this section. At the very bottom of the window are buttons for 'View Problems...', 'Protein Reports...', 'Ramachandran Plot...', and a 'Reset' button.

# Protein Preparation Wizard in Schrodinger

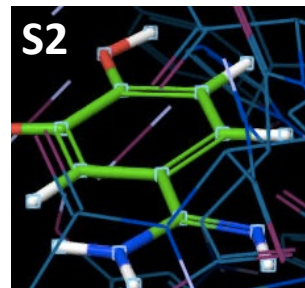
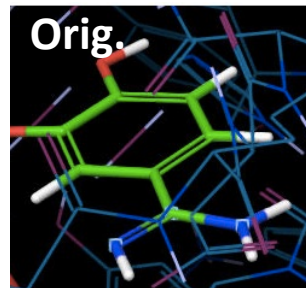
- **Import structure**
  - From RCSB website:
    - Diffraction data: for refining data with Primax
    - Biological unit: merge into a single entry
  - From local PDB files
- **Preprocess options**
  - Align one protein to another protein
  - Correct metal ionization states to ensure proper formal charge and force field treatment
  - Add sulfur bond between sulfur atoms that are within 3.2 Å of each other
  - Convert selenomethionines to methionine
  - Protein refinement with Prime
  - Cap protein termini with ACE and NMA residues
  - Remove water molecules at the user's discretion



# Protein Preparation Wizard in Schrodinger

## Review and Modify Tab

- **Analyze workspace**
- Delete waters (bulk water, water away from binding site, ...)
- Correct the ionization and tautomeric states of listed HET groups
  - Generate States: run an epic job at the target PH range



- Display state penalty

Protein Preparation Wizard

Job prefix: prepwizard Host: localhost (20)

Display hydrogens: ☐ None ☐ Polar only ☒ All ligand, polar receptor ☐ All

Import and Process Review and Modify Refine

Analyze Workspace

☒ Fit on select ☐ Display selection only ☐ Pick

Select Hets/Waters within 5.0 Å of selected chains

Select Lone Waters ☐ Invert Selection ☐

Chain Name	Water No.	Chain	Residue No.
A	1	A	602
	2	A	604
	3	A	605
	4	A	606

Het No.	Het Name	Orig.	S2	S3
1	A:CA (507)	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
2	A:CL (508)	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
3	A:Z34 (500)	<input type="checkbox"/>	<input checked="" type="checkbox"/>	<input type="checkbox"/>

Regenerate States

pH: 7.0 +/- 2.0

View Problems... Protein Reports... Ramachandran Plot...

State penalty: 2.63 kcal/mol; H-Bond count: 0; Q: 0

Reset



# Protein Preparation Wizard in Schrodinger

## Refine Tab

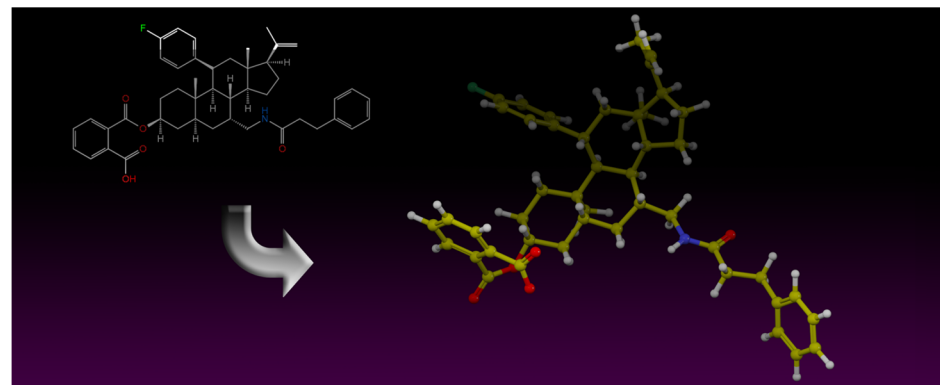
- **Optimizing the H-bonding network by**
  - reorienting water, amide groups, imidazole ring, ...
  - Use crystal symmetry: important when part of structure is present in the asymmetric unit
  - Two options for pH
    - PROPKA
    - simplified rules
      - very low: protonate ASU, GLU, HIS
      - low: protonate HIS
      - neutral: normal biological state
      - high: deprotonate cystines
  - Optimize H-bond interactively
- **Remove waters with less than a specified number of H-bond**
- **Restrained minimization**

The screenshot shows the 'Protein Preparation Wizard' window, specifically the 'Refine' tab. The 'Job prefix' is 'prepwizard' and the 'Host' is 'localhost (20)'. Under 'Display hydrogens', 'All ligand, polar receptor' is selected. The 'H-bond assignment' section has 'Sample water orientations' checked, 'Use crystal symmetry' and 'Minimize hydrogens of altered species' unchecked, 'Use PROPKA' selected with pH '7.0', and 'Label pKas' unchecked. Under 'Use simplified rules', 'Neutral' is selected. The 'Optimize' button is highlighted with a red box. Below it is the 'Interactive Optimizer...' button. The 'Remove waters' section has 'Beyond hets' checked with a value of '3.0' Å, and 'With fewer than' set to '3' H-bonds to non-waters. The 'Restrained minimization' section has 'Converge heavy atoms to RMSD' set to '0.30' Å, 'Hydrogens only' unchecked, and 'Force field' set to 'OPLS3e'. The 'Minimize' button is highlighted with a red box. At the bottom, there are buttons for 'View Problems...', 'Protein Reports...', and 'Ramachandran Plot...'. A status message reads 'Restrained minimization job incorporated.' and a 'Reset' button is in the bottom right corner.

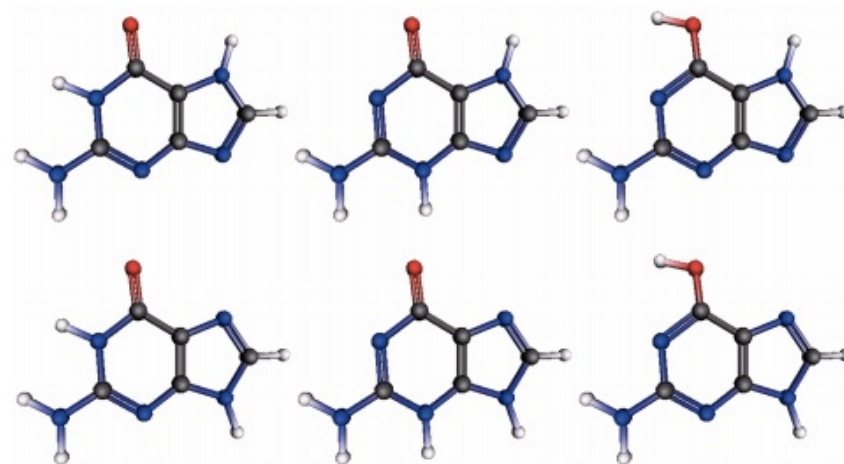
# Ligand Preparation

- Take 2D or 3D structures and produce low energy 3D structures
- Generate reasonable atomic coordinates for a ligand dataset

- tautomeric states
- ionization states
- ring conformations
- stereoisomers
- conformers



<https://www.schrodinger.com/ligprep>



Generation of multiple tautomeric forms  
of the ring system in a guanine ligand

*J. Chem. Inf. Model.* 2009, 49, 6, 1535–1546

# Ligand Preparation with LigPrep

- **Import structures:** from project, SD, SMILES format ....
- **Filter criteria:**
  - properties
  - general attributes: MW, number of atoms, ...
  - functional group counts
- **Force field**
- **Generate ionization states:**
  - Ionizer
  - Epik (recommended)
- **Desalt:** removes extra water molecule or counter ions that are present in ligand files that are originate from some structure databases
- **Generate tautomers:** keto-enol, sulfur/nitrogen, histidine, DNA base tautomerization
- **Stereoisomers**

LigPrep takes about 1-2 seconds on average to process a ligand.  
Result in difference in Epik State penalty (kcal/mol)

The screenshot shows the LigPrep software window with the following settings:

- Use structures from: Workspace (1 included entry)
- Filter criteria file: (empty field) [Create...] [Browse...]
- Force field: OPLS3e
- Ionization:
  - ☐ Do not change
  - ☐ Neutralize
  - ☒ Generate possible states at target pH: 7.0 +/- 2.0
- Using: ☐ Ionizer ☒ Epik
- ☐ Add metal binding states
- ☐ Include original state
- ☒ Desalt ☒ Generate tautomers
- Stereoisomers:
  - Computation:
    - ☒ Retain specified chiralities (vary other chiral centers)
    - ☐ Determine chiralities from 3D structure
    - ☐ Generate all combinations
  - Generate at most: 32 per ligand
  - ☐ For SD V2000 input, generate enantiomers if the chiral flag is 0
- Output format: ☒ Maestro ☐ SDF

Job name: ligprep\_1 [Run]

Host=ada-cpu-48, Incorporate=Append new entries as a new group

# Hands-on Session 1

## Structure Visualization and Preparation with Maestro

1. Creating Projects and Importing Structures
2. Preparing Protein Structures (**Protein Preparation Wizard**)
3. Preparing Ligand Structures (**LigPrep**)
4. Visualizing Protein-Ligand Complexes (configuration bar, Ligand Interaction Diagram)

# Steps of structure based virtual screening

explore poses of  
ligand in the  
binding site

**Docking**

- Rigid receptor docking
- Induced fit docking
- Covalent docking
- ...

quantify the poses  
with a function

**Scoring**

- Docking Score
- Glide Score
- Emodel
- ...

improve poses  
and select  
compounds

**Refining**

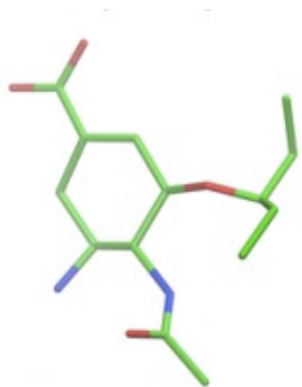
- RMSD
- Enrichment
- Receiver operator characteristic plots
- ...



# Docking fits ligands to a rigid receptor in a pose

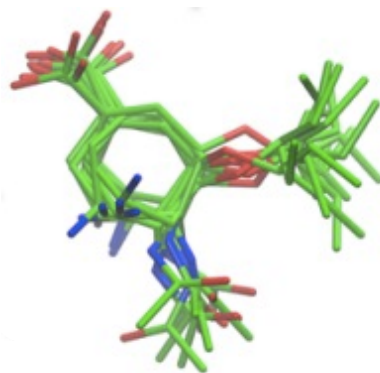
## Docking

Search for the best-scoring binding pose for a given ligand  
Rigid receptor docking with Glide HTVS, SP, XP  
Receptor is rigid  
Ligand is flexible



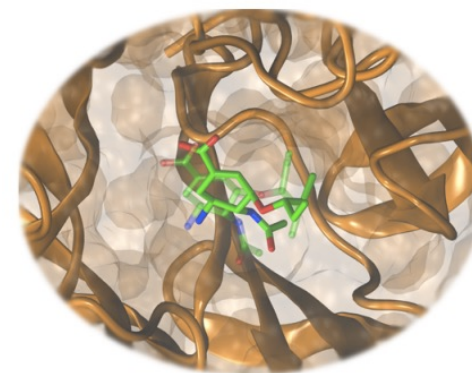
prepared  
ligand

Ligand sampling



collection of ligand  
conformations

docking

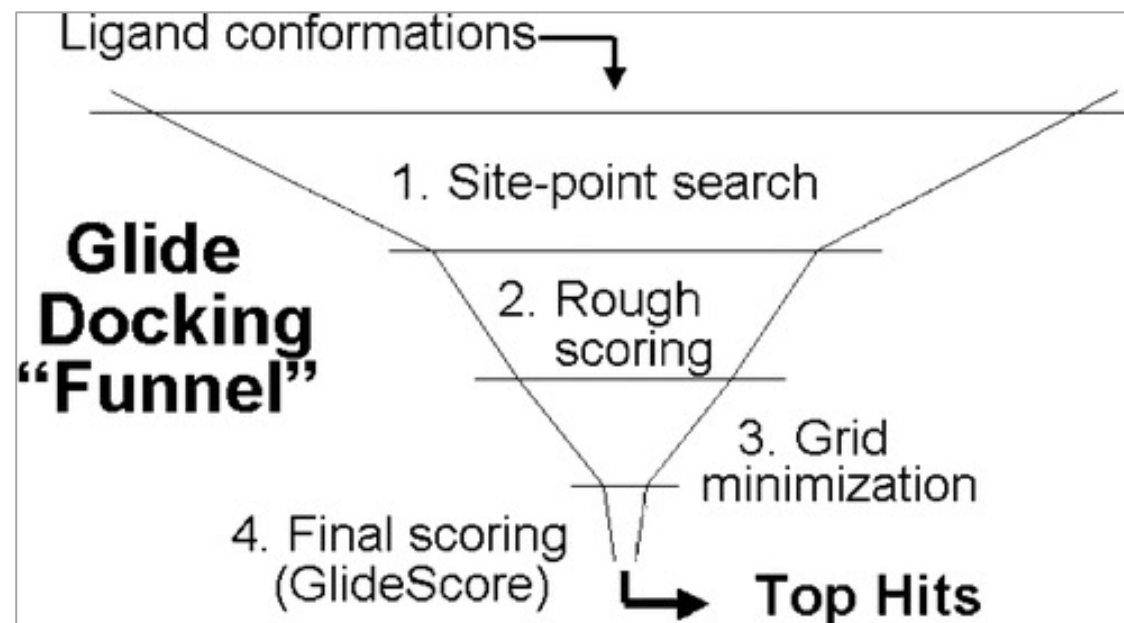


ensemble of  
receptor-ligand poses

# Ligand Docking

- Procedure
  - Prepare the protein
    - Missing atoms/side chains
    - Protonation state
    - Flexible side chains
  - Prepare the ligand
    - Protonation state
  - Create a docking grid
    - Specify where to dock the ligand
  - Dock the ligand(s)
  - Scoring
  - Refinement

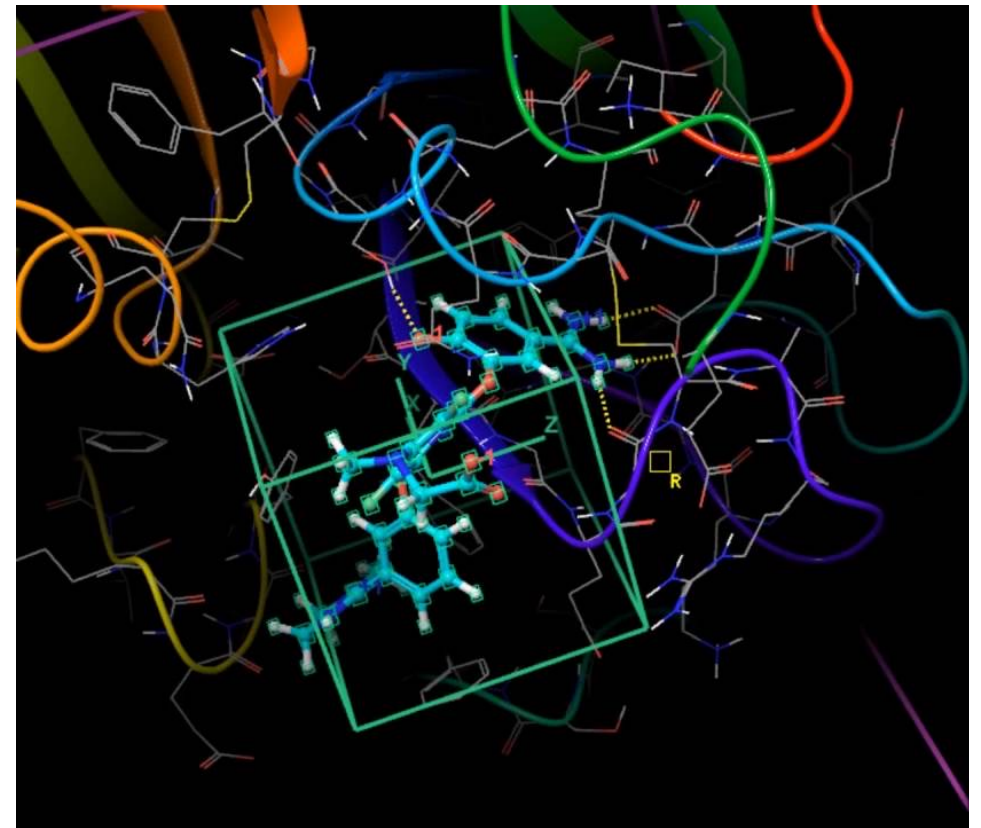
- Glide docking hierarchy



Glide docking "funnel", showing the Glide docking hierarchy.

# Binding Pocket – Grid Generation

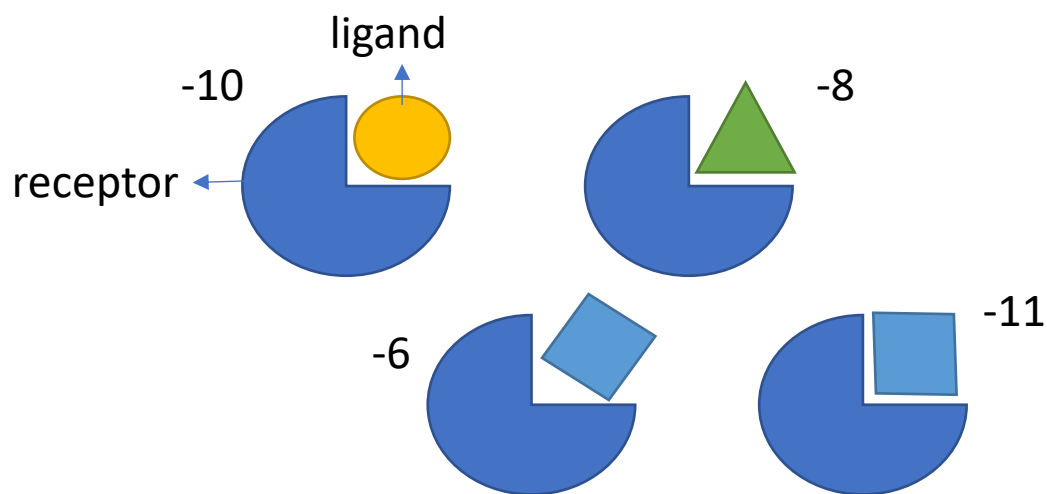
- Utilities to suggest binding sites – such as Schrödinger's SiteMap
- Use binding site from crystal structures with a bound ligand (cognate ligand)
- Binding Pocket Grid
  - Bounding box where docking is performed
  - Too small
    - ligands won't dock
    - miss good ligands
  - Too big
    - increase computational cost substantially
    - miss good binding poses
- Is the binding pocket rigid or flexible?
  - Molecular dynamics simulations can be used to investigate the stability of the binding pocket



# Steps of structure based virtual screening



A scoring function very **roughly** approximates the **binding affinity** of a ligand to a protein given a binding pose.



## Illustration of binding pose ensembles

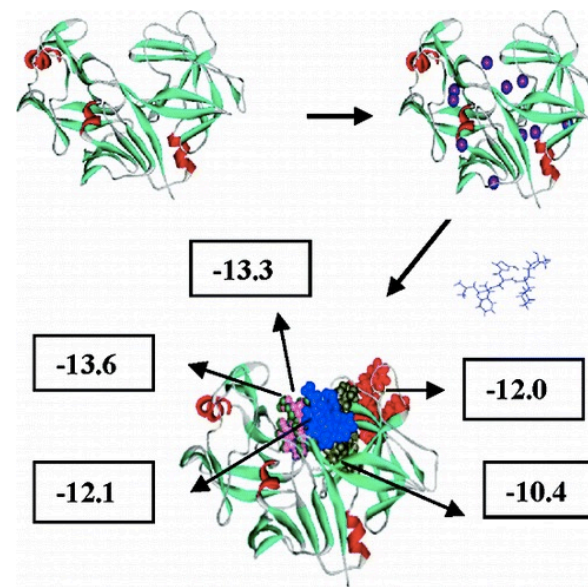


Figure from *J. Chem. Inf. Model.* 2011, 51, 10, 2515–2527

# Scoring evaluates the ligand pose



- Do not correlate with  $IC_{50}$ ,  $K_d$ ,  $EC_{50}$ , etc
- More negative the score, the better
- Are optimized to give good enrichment
  - Separate good from bad ligands
  - Limit the number of ligands that need to be investigated further



# GlideScore and Emodel

- Glidescore

rank-order compounds to separate compounds that bind strongly (actives) from those that don't (inactives)

Scoring Function	Computing Time	When to Use
SP	5 – 20 sec/molecule	<u>First pass</u> virtual screening on large databases/hit generation
XP	3-5 min/molecule	<u>Refinement</u> of a smaller dataset for lead optimization

- SP seeks to minimize false negatives while XP seeks to minimize false positives
- The XP scoring function includes more stringent terms for modeling desolvation, hydrophobic effects, and charged interactions

- Emodel

- primarily defined by protein-ligand coulomb-vdW energy with a small contribution from GlideScore
- Choose the best-docked structure for each ligand

# Glide Docking SP

$$\text{GScore} = 0.05 * \text{vdW} + 0.15 * \text{Coul} + \text{Lipo} + \text{Hbond} + \text{Metal} + \text{Rewards} + \text{RotB} + \text{Site}$$

Components	Description
VdW	Van der Waals energy. This term is calculated with reduced net ionic charges on groups with formal charges, such as metals, carboxylates, and guanidiniums.
Coul	Coulomb energy. This term is calculated with reduced net ionic charges on groups with formal charges, such as metals, carboxylates, and guanidiniums.
Lipo	Lipophilic term, which is a pairwise term in SP but is derived from the hydrophobic grid potential for XP. Rewards favorable hydrophobic interactions.
HBond	Hydrogen-bonding term. This term is separated into differently weighted components that depend on whether the donor and acceptor are neutral, one is neutral and the other is charged, or both are charged.
Metal	Metal-binding term. Only the interactions with anionic or highly polar acceptor atoms are included. If the net metal charge in the apo protein is positive, the preference for anionic or polar ligands is included; if the net charge is zero, the preference is suppressed.
Rewards	Rewards and penalties for various features, such as buried polar groups, hydrophobic enclosure, correlated hydrogen bonds, amide twists, and so on. This category covers all terms other than those explicitly mentioned.
RotB	Penalty for freezing rotatable bonds.
Site	Polar interactions in the active site. Polar but non-hydrogen-bonding atoms in a hydrophobic region are rewarded.

# Glide Docking XP (Extra Precision)

- Increase computational cost
- Glide SP with additional Extra Precision terms
- Anchor fragments of the docked ligand, typically rings, are chosen from the set of SP poses and the molecule is re-grown bond by bond from these anchor positions
- Rewards occupancy of well-defined hydrophobic pockets by hydrophobic ligand groups which is often under-estimated
- Includes improvements to the scoring of hydrogen bonds as well as detection of buried polar groups, and detection of pi-cation and pi-pi stacking interactions

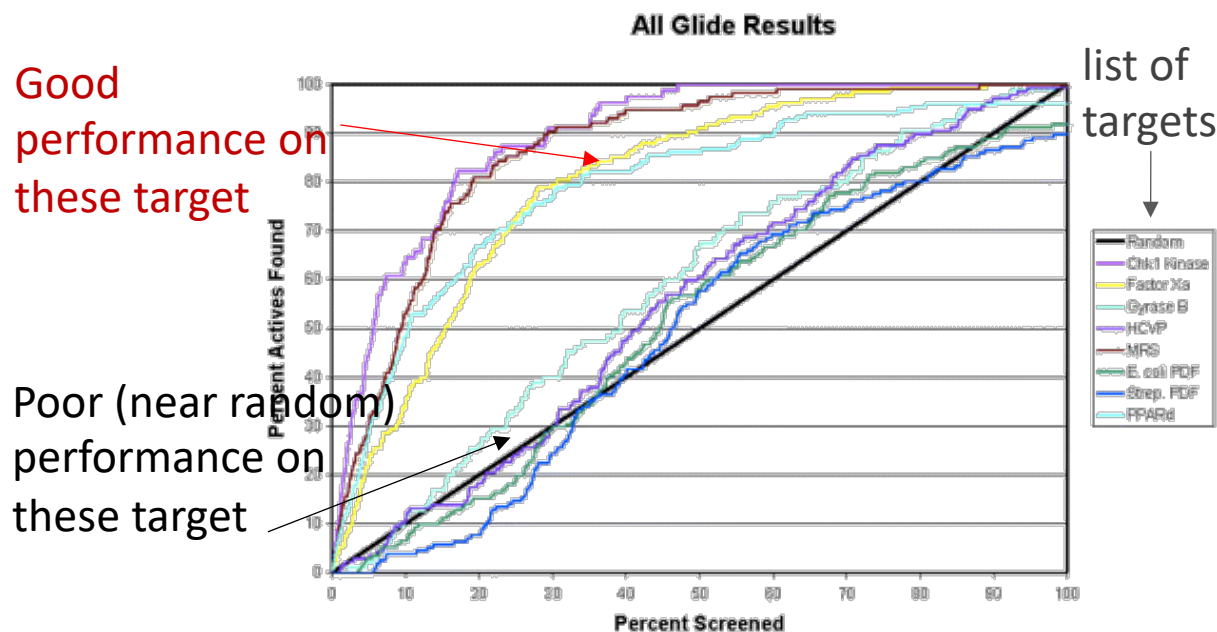
# Filtering refines the ligand evaluation

Docking

Scoring

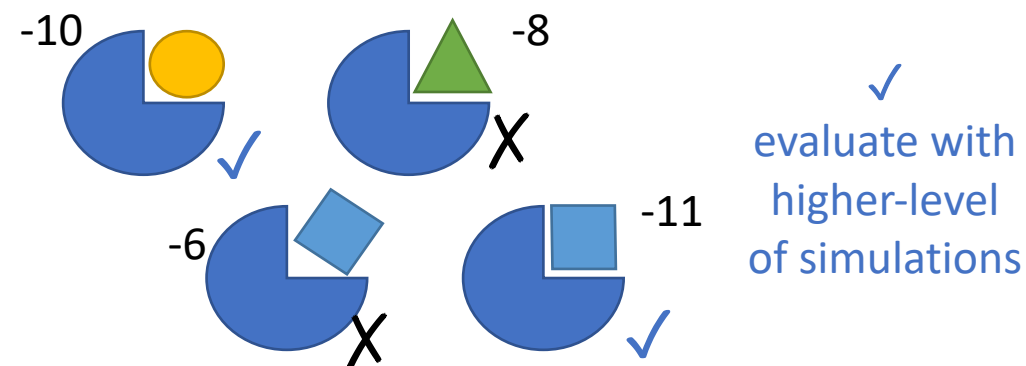
Refining

- Docking performance



J. Med. Chem. 2006, 49, 20, 5912–5931

- Screening compounds for further evaluation



# Hands-on Session 2

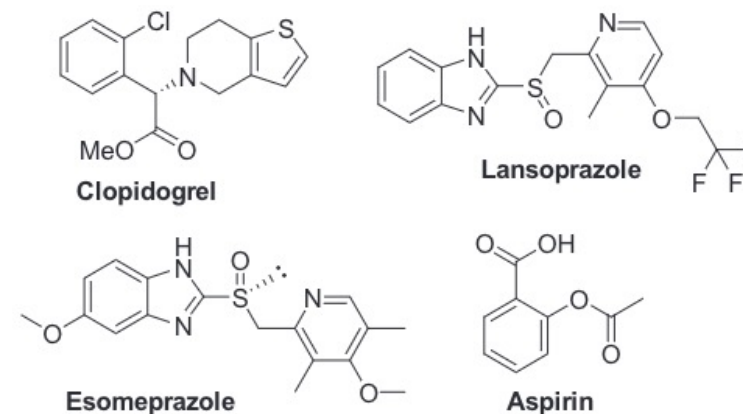
## Structure-Based Virtual Screening Using Glide

1. Virtual Screening Prerequisites
2. Importing Structures
3. Generating a Receptor Grid
4. Docking the Cognate Ligand and Screening Compounds
5. Analyzing Results and Binding-Site Characterization

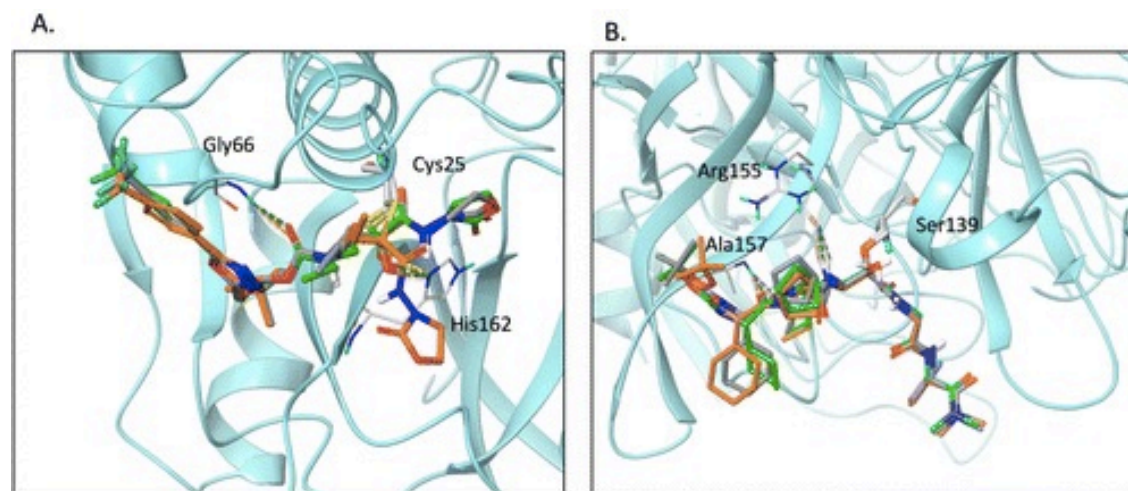


# Covalent Docking

- Nearly 30% of the marketed drugs targeting enzymes known to act by covalent inhibition
- The inhibition can be either reversible or irreversible
- Covalent inhibitors derive their activity not only from the formation of a covalent bond between the target and the ligand but also from stabilizing non-covalent forces in the binding pocket



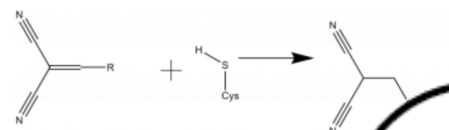
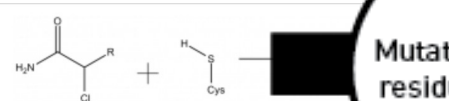
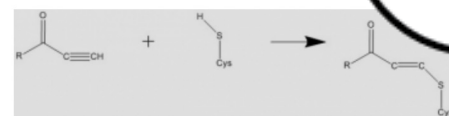
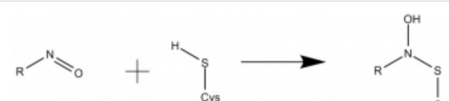
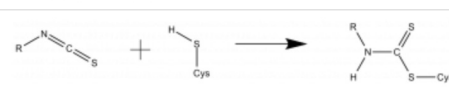
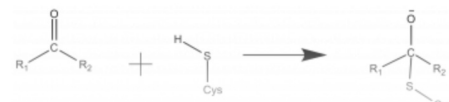

Examples of drug act through covalent mechanisms.



Examples of covalent complexes. A) Cathepsin K structure (PDB ID 1YT7) with the cocystal ligand, B) HCV NS3 protease structure (PDB ID 2F9U) with the cocystal ligand.

# Covalent Docking

Custom Covalent Reactions

Index	Reaction	Ligand SMARTS	Receptor SMARTS	CDOCK File
1		<chem>[C,c]=[C,c]-[C,c]#[N,n]</chem>	<chem>[C]-[S,O;H1,-1]</chem>	<a href="#">Download</a>
2		<chem>[C,c]([C])=[O]</chem>	<chem>[C]-[S,O;H1,-1]</chem>	<a href="#">Download</a>
3		<chem>[C,c]-[C,c,S,s]=[O]</chem>	<chem>[C]-[S,O;H1,-1]</chem>	<a href="#">Download</a>
4		<chem>[N]=[O,S]</chem>	<chem>[C,c]-[S,O;H1,-1]</chem>	<a href="#">Download</a>
5		<chem>[N]=[C]=[S]</chem>	<chem>[#6]-[S;H1]</chem>	<a href="#">Download</a>
6		<chem>[C,c]=[O,S]</chem>	<chem>[C,c]-[S,O;H1,-1]</chem>	<a href="#">Download</a>
7		<chem>[C]=[C]-[C]=[O]</chem>	<chem>[C]([O])-[C]-[C]</chem>	<a href="#">Download</a>

Mutate reactive  
residue to ALA

Sample ligand  
conformations  
with ConfGen

Glide docking  
with positional  
constraint

Apply pose  
selection criteria

Covalently link  
ligand &  
attachment  
residue

Optimize  
ligand poses and  
residue geometry  
with Prime  
VSGB2.0

Rank poses  
using VSGB2.0  
and compute  
binding affinity

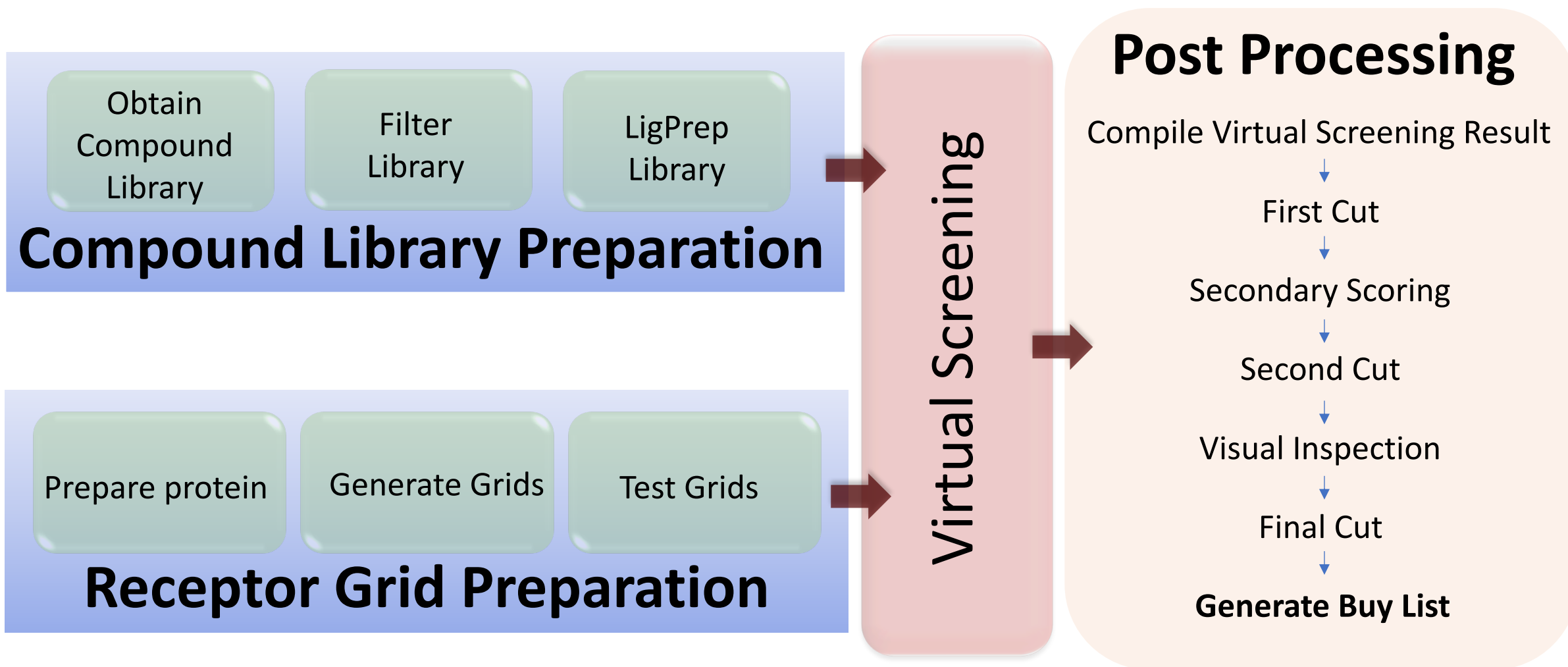
<https://www.schrodinger.com/newsletters/introducing-covdock-covalent-docking>

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# CovDock Uses Glide & Prime

- Main steps
  - Conventional non-covalent docking of pre-reactive species (**Glide**)
  - Formation of covalent attachment (via a number of different mechanisms)
  - Structural refinement of the covalent complex (**Prime**)
- Output: **cdock affinity, prime energy, ligand reaction site**
- Speed
  - Pose selection (default) protocol: 1~2 hour per ligand
  - Virtual screening protocol: 10x faster than default protocol
- Challenges for covalent docking
  - Bond formation, bond cleavage and bond rearrangements all require an explicit treatment of electronic degrees of freedom and, hence, a quantum mechanics (QM) approach.

# Drug Docking with Schrodinger: wrap-up



# Running Schrodinger on HPRC

## Schrödinger is a restricted software.

Usage of this software is restricted to subscribers of the [Laboratory for Molecular Simulation \(LMS\)](https://lms.hprc.tamu.edu/).

Running Schrodinger on Grace and Terra, please refer to: <https://hprc.tamu.edu/wiki/SW:Schrodinger>.

More about the Schrodinger: [documentation](#), [training](#)

## The LMS also holds license for:

- Discovery Studio
- MOE
- Amber
- Material Studio
- Gaussian
- ADF
- Molpro
- Chemissian
- NBO
- AIMALL Professional

# Need Help? Contact the HPRC Helpdesk

Website: [hprc.tamu.edu](https://hprc.tamu.edu)

Email: [help@hprc.tamu.edu](mailto:help@hprc.tamu.edu)

Telephone: (979) 845-0219

## Help us, help you -- we need more info

- Which Cluster (Terra, Grace)
- NetID (NOT your UIN)
- Job id(s) if any
- Location of your jobfile, input/output files
- Application used if any
- Module(s) loaded if any
- Error messages
- Steps you have taken, so we can reproduce the problem